

LIGNANS OF *PIPER CLUSII**

S. K. KOUL, S. C. TANEJA, K. L. DHAR and C. K. ATAL

Regional Research Laboratory, Jammu 180001, India

(Revised received 11 September 1982)

Key Word Index—*Piper clusii*; Piperaceae; lignans; (–)-hinokinin; (–)-cubebin; (–)-deoxypodorhizon; (–)-dihydrocubebin; (–)-clusin; asaronaldehyde; ^{13}C NMR; CD.

Abstract—From the petrol extract of *Piper clusii* five lignans were isolated. One of the lignans (–)-clusin is assigned the structure (–)-2-furanol-4(1,3-benzodioxol-5-ylmethyl) tetrahydro-3(3,4,5-trimethoxyphenyl) methyl. This is the first report of this compound from a natural source. Asaronaldehyde and sitosterol were also present.

INTRODUCTION

Piper clusii is reported to have some medicinal properties [1]. Previous work reports the presence of piperine and sesamin as the only lignans from the leaves of the plant [2]. The present investigations on this plant have proved it to be a rich source of lignans, viz. (–)-hinokinin, (–)-cubebin, (–)-dihydrocubebin and (–)-deoxypodorhizon. One of the lignans named (–)-clusin (**1**) is being reported for the first time. It has been assigned the structure (–)-2-furanol-4(1,3-benzodioxol-5-ylmethyl) tetrahydro-3(3,4,5-trimethoxyphenyl)methyl. The presence of sitosterol and asaronaldehyde is also reported.

RESULTS AND DISCUSSION

Chromatographic separation of the extract (benzene-ethyl acetate) resulted in the isolation of compound **1** (viscous mass). Elemental analysis gave a molecular formula of $\text{C}_{22}\text{H}_{26}\text{O}_7$, M^+ m/z 402. IR gave a strong absorption at 3400 cm^{-1} indicating the presence of a hydroxyl function. ^1H NMR in CDCl_3 showed a sharp singlet at δ 5.86 for two protons assigned to the methylenedioxy group. Signals for three methoxyl groups were observed at δ 3.90. Signals for benzylic protons and methine protons were observed as an envelope between δ 2.20 and 2.80. A broad singlet at δ 5.10 is ascribed to a

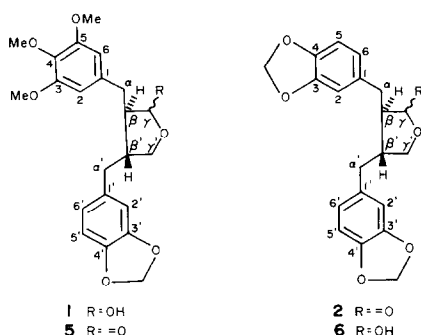
hemiacetalic proton. Aromatic protons were located between δ 6.20 and 6.70. The $-\text{O}-\text{CH}_2-$ protons were observed from δ 3.9 to 4.1 as a multiplet.

Acetylation gave a monoacetate (**1a**), a semi-solid, which analysed for $\text{C}_{24}\text{H}_{28}\text{O}_8$. The ^1H NMR, showed a signal for the hemiacetal C-1 shifted to δ 6.10 whereas the signal at δ 5.10 disappeared. The other signals remained unchanged.

Oxidation with Collin's reagent gave one product which analysed for $\text{C}_{22}\text{H}_{24}\text{O}_7$. IR showed a strong signal at 1770 cm^{-1} indicative of a γ -lactone while the band at 3460 cm^{-1} (observed in **1**) disappeared. The ^1H NMR (CDCl_3) showed a four proton envelope at δ 2.43 and a narrow two proton envelope at δ 2.70 assigned to benzylic and β, β' -protons, respectively. This situation of the two envelopes arises when the stereochemistry at β and β' is *trans* [10]. Moreover, a multiplet centred at δ 4.00 also supports the *trans* stereochemistry at β and β' . In the case of *cis*- β, β' the signals for benzylic and methine protons are generally observed as a multiplet between δ 2.1 and 3.3. Other signals remained practically unchanged. The spectral data of the compound, including the mass spectral fragmentation, are in agreement with deoxypodorhizon [7]. An additional proof of the stereochemistry at β and β' was obtained by comparing the CD curve of the lactone with that of (–)-hinokinin [3]. Both show a negative Cotton effect. This further confirms that the stereochemistry at β and β' in the parent compound (**1**) is also *trans*.

A compound with a similar structure to **1** has also been synthesized by Kiyoshi *et al* [4] as an intermediate during the synthesis of the anticancer lactone (+)-steganacin. However, Tomioka's compound is dextrarotatory with $[\alpha]_D + 48.1$ (EtOH) while **1** is laevorotatory with $[\alpha]_D - 34.4$ (CHCl_3). Therefore, clusin appears to be the enantiomer of the compound synthesized by Kiyoshi *et al*. The structure is also supported by its mass spectrum.

^{13}C NMR proved to be invaluable in confirming the structure of **1**. A comparison of the ^{13}C NMR spectrum of (–)-cubebin (**6**) with that of **1** and its lactone (**5**) is presented in Table 1. The values at the β and β' -carbons are in complete accordance with that of (–)-cubebin. The values of the hemiacetal carbons observed at δ 103.1 are also close to those in cubebin (δ 103.5). The other values are also similar and further support the proposed structure of (–)-clusin.



*Communication No. RRL/81-025.

Table 1. ^{13}C chemical shift values of 1, 5 and 6

Carbon position	1	5	6
α	39.0	35.37	39.5
α'	38.7	38.5	38.7
β	51.5	41.42	53.1
β'	45.1	46.7	45.1
γ	103.1	178.0	103.5
γ'	73.9	71.28	72.7
1	130.7	129.24	132.3
1'	133.9	133.9	133.9
2	105.9	107.2	108.3
6	105.9	107.2	121.9
2'	108.0	108.0	109.3
5'	108.8	108.5	109.0
6'	121.1	121.95	121.6
3	153.1	153.7	147.5
5	153.1	153.77	109.5
4	145.9	146.8	145.9
3'	148.3	147.07	147.5
4'	147.5	147.37	145.9
O-CH ₂ O	101.1	101.2	101.1
OMe	56.3	56.3	—

EXPERIMENTAL

All mps determined are uncorr. IR spectra were recorded in KBr pellets. ^1H NMR at 60 MHz in CDCl_3 and MS at 70 eV. Specific rotations were determined in CHCl_3 soln.

Dried plant material of *P. elusii* Cass DC. (200 g) was extracted with petrol (60–80°) in a Soxhlet. Removal of solvent yielded a resinous mass (16 g) which on repeated CC over Si gel and elution with C_6H_6 and C_6H_6 -EtOAc in increasing proportions furnished seven isolates in pure form designated 1–7.

1. Gummy solid, M^+ m/z 402, analysed for $\text{C}_{22}\text{H}_{26}\text{O}_7$ (requires C, 65.65; H, 6.52; observed C, 65.39; H, 6.51 %); $[\alpha]_{\text{D}}^{26} - 34.5^\circ$ (CHCl_3 ; c 1.0). IR prominent bands at 1590 ($\text{C}=\text{C}$), 3400 ($-\text{OH}$), 928 cm^{-1} ($-\text{OCH}_2\text{O}$); other bands at 1487, 1440, 1320, 1230, 1120 and 1020 cm^{-1} . ^1H NMR: envelope between δ 2.20 and 2.80 (6H, four benzylic and two methine protons), s at 3.90 (9H, aromatic OMe), 5.86 (2H, s , $-\text{OCH}_2\text{O}$) and m 6.20–6.70 (five aromatic protons), m 3.9–4.1 (2H, $\text{O}-\text{CH}_2-$) and br s at 5.10 (1H) for a hemiacetal proton. The signal at δ 4.30 was exchangeable with D_2O . MS m/z 402 (base peak), 385, 384, 249, 182, 181, 127, 121 and 119. Acetylation with pyridine- Ac_2O gave the monoacetate (1a), a semi-solid, M^+ m/z 444, analysed for $\text{C}_{24}\text{H}_{28}\text{O}_8$. IR showed disappearance of the band at 3400 and

O

||

appearance of two bands at 1730 and 1245 cm^{-1} ($-\text{O}-\text{C}-\text{Me}$). ^1H NMR showed a downfield shift of the hemiacetalic proton observed at δ 6.10 (1H) while the other signals were observed at δ 3.89 (9H, aromatic OMe), 5.90 (2H, s , $-\text{OCH}_2\text{O}$), 6.21–6.73 (m , five-aromatic protons), 3.9–4.16 (m , $-\text{OCH}_2$) and 2.23 and 2.86 as an envelope (6H, four benzylic and two methine protons). MS m/z 444, 402, 385, 384, 249, 182, 181, 121, 119 and 117.

Oxidation of 1 with Collin's reagent gave a gummy mass, analysed $\text{C}_{22}\text{H}_{24}\text{O}_7$, M^+ m/z 400.4125 (requires C, 65.99; H, 6.04; observed C, 66.12; H, 6.12 %); $[\alpha]_{\text{D}}^{26} - 11.1^\circ$ (CHCl_3 ; c 0.72) negative Cotton effect at 280 nm. From the spectral data the compound was identified as (–)-deoxypodorhizon in comparison with an authentic sample [7], (co-TLC, superimposable IR).

2. Semisolid analysed for $\text{C}_{29}\text{H}_{18}\text{O}_6$, M^+ at m/z 354 (requires

C, 67.69; H, 5.12; observed C, 68.00; H, 5.08 %); $[\alpha]_{\text{D}}^{26} - 19.1^\circ$. IR bands at 1772, 1600 and 923 cm^{-1} . ^1H NMR signals: δ 2.43 (4H, m , $\phi\text{-CH}_2$), 3.9 (envelope, 2H, OCH_2), 2.83 (envelope, 2H, $-\text{CH}$), 5.86 (1H, s , $-\text{OCH}_2\text{O}$) and 6.5 (6H, m , Ar-H). The compound was identified as (–)-hinokinin by comparison with an authentic sample [5] (co-TLC, superimposable IR).

3. Crystals from EtOH, mp 114–115°. The compound was identified as asaronaldehyde by comparison with an authentic (lit. [6] mp 114°) (co-TLC and superimposable IR).

4. Mp 137°, analysed for $\text{C}_{29}\text{H}_{20}\text{O}$, M^+ at m/z 414. The compound was identified as sitosterol by comparison with an authentic sample (lit. [6] mp 137°) (co-TLC and superimposable IR).

5. Semisolid, analysed for $\text{C}_{22}\text{H}_{24}\text{O}_7$, M^+ at m/z 400 (requires C, 65.99; H, 6.04; observed C, 65.83; H, 6.13 %); $[\alpha]_{\text{D}}^{26} - 26.1^\circ$. From the spectral data (IR, ^1H NMR and MS) the compound was identified as (–)-deoxypodorhizon by comparison with an authentic sample [7], (Co-TLC and superimposable IR).

6. Mp 131.5°, analysed for $\text{C}_{20}\text{H}_{20}\text{O}_6$, M^+ at m/z 356 (requires C, 67.40; H, 5.65; observed C, 67.49; H, 5.59 %). IR bands at 3330, 1605 and 930 cm^{-1} . ^1H NMR signals δ 2.66 (envelope, 2H, $\phi\text{-CH}_2$), 3.96 (envelope, 2H, $\text{O}-\text{CH}_2-$), 5.26 (1H, br s , $-\text{CH}$), 5.96 (4H, s , $-\text{OCH}_2\text{O}$) and 6.6 (6H, m , Ar-H). Monoacetate analysed for $\text{C}_{22}\text{H}_{22}\text{O}_7$, M^+ m/z 398 (requires C, 66.32; H, 5.56; observed C, 66.39; H, 5.59 %). ^1H NMR signals: δ 2.0 (3H, s , $-\text{OCOMe}$), 3.8 (envelope, 2H, $-\text{OCH}_2$), 6.1 (1H, s , $-\text{CH OAc}$). The compound was identified as (–)-cubebin by comparison with an authentic sample (lit. [8] mp 131°) (Co-TLC and superimposable IR).

7. Mp 102°, analysed for $\text{C}_{20}\text{H}_{22}\text{O}_6$, M^+ at m/z 358 (requires C, 67.21; H, 6.20; observed C, 67.38; H, 6.23 %); $[\alpha]_{\text{D}}^{26} - 32.1^\circ$. IR bands at 3310, 1601 and 924 cm^{-1} . ^1H NMR signals: δ 2.63 (4H, d , $J = 7$ Hz, $\phi\text{-CH}_2$), 3.56 (envelope 4H, $-\text{OCH}_2$), 4.03 (2H, br s , exchangeable with D_2O), 5.93 (4H, s , $-\text{OCH}_2\text{O}$) and 6.63 (6H, m , Ar-H). Diacetate (gummy solid), ^1H NMR signals for methyleneoxy and methine proton were shifted to δ 4.06 and 3.80, respectively, and the other signals remained essentially unchanged. MS fragments at m/z 442, 382, 322, 187 (base peak), 186, 174, 136 and 135. 7 was identified as (–)-dihydrocubebin (lit. [9] mp 101–102°) (Co-TLC, mmp and superimposable IR).

Acknowledgements—We are thankful to C.D.R.I., Lucknow for the CD and to the Instrumentation Division, RRI., Jammu for spectral data.

REFERENCES

- Kirtikar, K. R. and Basu, B. D. (1961) *Indian Medicinal Plants* p. 2127. Mohan Basu, Allahabad.
- Hansel, R. and Zander, D. (1961) *Arch. Pharm.* **294**, 699.
- Burden, R. S., Crombie, L. and Whiting, D. A. (1969) *J. Chem. Soc. C* 693.
- Kiyoshi, T., Ishiguro, T. and Koga, K. (1980) *Tetrahedron Letters* **21**, 2973.
- Rao, C. B. S. (1978) *Chemistry of Lignans* pp. 107–108. Andhra University Press, Waltair.
- (1965) *Dictionary of Organic Compounds* pp. 3155 and 2902. Fyre & Spottiswoode, London.
- McDoniel, P. B. and Cole, J. R. (1972) *J. Pharm. Sci.* **61**, 1992.
- Batterbee, J. E., Burden, R. S., Crombie, L. and Whiting, D. A. (1969) *J. Chem. Soc.* 2470.
- Tillekeratne, L. M. V., Jayamanee, D. T., Weerasuria, K. D. V. and Gunatilaka, L. (1982) *Phytochemistry* **2**, 476.
- Corrie, J. E. T., Green, G. H., Ritchie, E. and Taylor, W. C. (1970) *Aust. J. Chem.* **23**, 133.